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Amplidiag® Multiplex PCR Master Mix (2x)

High-performance
ready-to-use
hot-start master mix

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Amplidiag® Multiplex PCR Master Mix (2x) is concentrated high-performance ready-to-use master mix. The master mix has been optimized for **high sensitivity and specificity** in both **multiplex** and **singleplex real-time PCR** and **PCR assays**.

Amplidiag® Multiplex PCR Master Mix (2x) has extremely low level of residual DNA, making it an unsurpassed solution for **challenging applications**. The Master Mix comes with Mobidiag's proprietary Taq DNA polymerase and utilizes **antibody-based hot-start technology** with very low residual activity before the initial denaturation step, thus increasing the specificity of the PCR.

Maximum multiplexing capability and high efficiency

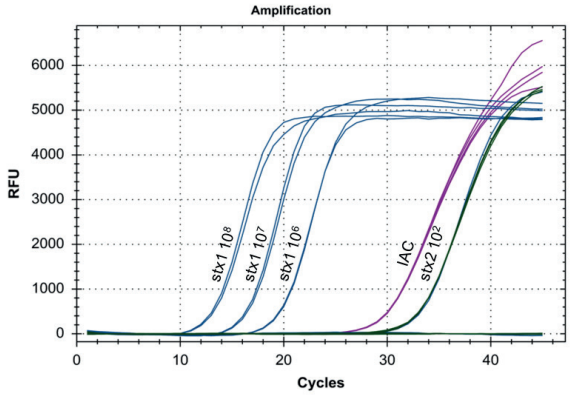
1. Efficient amplification with high sensitivity of low abundance targets in multiplex assays
2. Efficient amplification and ultra-sensitive detection in inhibitory sample materials
3. Extremely low content of residual DNA makes the master mix ideal for detection of bacterial DNA in multiplex assays

Amplidiag® Multiplex PCR Master Mix (2x)

Product no.	Package size
AD-MM103x2	2x1,1ml
AD-MM103x10	10x1,1ml

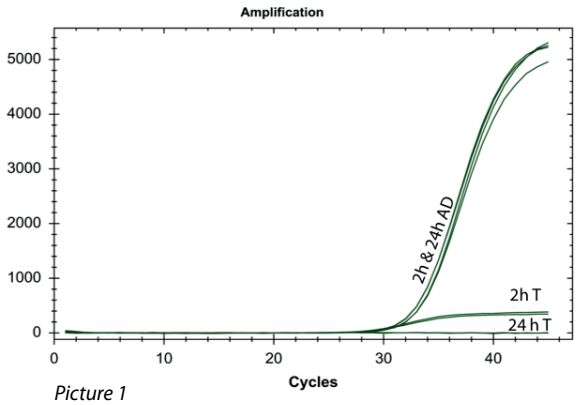
For research use only

No Cq lag in multiplex assays



One hundred copies of Shiga toxin type 2 (stx2) and 1000 copies of Internal Amplification Control (IAC) amplicons were amplified in the presence of three Shiga toxin type 1 (stx1) amplicon amounts: 10^6, 10^7 or 10^8 copies. As a comparison, all three different stx1 amplicon amounts, stx2 and IAC were each run separately. The combined results show invariant amplification, i.e. no Cq lag, for the different targets (stx1, stx2, IAC) in multiplex settings when compared to single plex runs. The labels used in the set up were: FAM (stx1), HEX (stx2) and Cy5 (IAC).

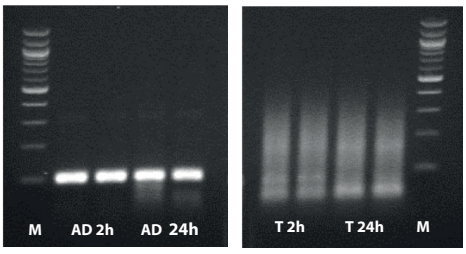
High performance even after 24 h in RT



Picture 1

Reaction setups were made with either Amplidiag® Multiplex PCR Master mix or a non-hot start Taq enzyme. The complete reactions were incubated either 2h or 24h at ambient temperature before run.

Picture 1 shows amplification results (in duplicate) after incubations. The Cq values and end point intensities in the Amplidiag® Multiplex PCR Master mix reactions (AD) were not affected by the incubation times, whereas the amplifications in reactions with non-hot start Taq enzyme (T) were severely deteriorated.



Picture 2

Picture 2 shows a gel run (in duplicate) of the incubated reactions after amplification: Amplidiag® Multiplex PCR Master mix (AD) reactions on left and non-hot start Taq enzyme (T) reactions on right. M = Marker. There is no signal loss for the 100 bp target in AD reactions whereas the non-hot start enzyme reactions fail in amplification.

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